



EFFECTS OF TURMERIC OIL AS A DIETARY SUPPLEMENTS ON THE HAEMATOLOGY AND SERUM BIOCHEMICAL INDICES OF BROILER CHICKENS

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Abstract

The objective of this current study was to examine the effects of turmeric oil as a dietary supplement on some hematological and serum biochemical indices of broiler chicken. A total of two hundred 1-day-old broiler chicks (Ross 308 strain) were randomly distributed to five treatments of 4 replicates consisting of 10 birds each in a completely randomized design. Birds in treatment 1 (T1) were fed basal diet + 0 % turmeric oil (TOL), T2, T3, T4 and T5 were fed basal diet supplemented with TOL at 0.1 %, 0.2 %, 0.3 % and 0.4 % respectively. Basal diet was formulated to meet the nutritional requirements of birds according to NRC (1994). Clean feed and water were also provided ad libitum throughout the experiment which lasted for 56 days. Results obtained showed that all the hematological parameters (Pack cell volume, hemoglobin, red blood cell, mean corpuscular volume, mean corpuscular hemoglobin, mean corpuscular hemoglobin concentration, white blood cells and its differentials) were significantly ($P < 0.05$) different among the treatments. Serum biochemical parameters (Total protein, albumin, globulin, cholesterol, alanine transaminase, aspartate transaminase) were influenced by the dietary supplementation of TOL ($P < 0.05$). Cholesterol level decreases as the dietary supplementation of TOL increases ($P < 0.05$). However, all values were within the normal physiological range for birds. It was concluded that TOL contains several bioactive chemicals which confers it the ability to perform multiple biological functions.

Keywords: *Broiler chickens, turmeric oil, hematology, serum biochemistry.*

INTRODUCTION

The increasing pressure of reducing the use of antibiotics as antimicrobial growth promoters for animals due to harmful residual toxicity effects of drugs observed in the food chain calls for alternative solutions to sustain the efficiency of current livestock production (Mahima et al., 2012; Oluwafemi et al., 2020). Among the potential alternatives is essential oil which has been found to be loaded with several bioactive chemicals, less toxic and free from residues (Henhxiao et al., 2018; Alagbe 2021). According to Franz and Novak (2009), essential oils are complex mixtures of volatile compounds produced by living organisms and isolated by physical means only (pressing and distillation) from the whole plant or plant part of known taxonomic origin. The composition of essential oil in plants depends on plant species and its age, harvesting periods, methods of processing, geographical location and soil types (Bouhaddouda et al., 2016; Oke et al., 2009; Hammer and Carson, 2011).

Scientific reports showed that essential oils (EOs) can perform several biological activities such as antibacterial, anti-inflammatory, cytotoxic, hypolipidemic, hepatoprotective, antiviral, miracidial and antioxidant (Michiels et al., 2009; Kim et al., 1995; Kommera et al., 2011; Giannenas et al., 2013). EOs are relatively cheap, effective, safe and perceived as growth promoters in poultry diets (Zhang et al., 2014), improve meat quality (Wenk, 2003, Alagbe and Akintayo, 2020), alteration of lymphocyte distribution in the gut (Purchiaroni et al., 2013; Oluwafemi et al., 2020), potentiate the immune response (Li et al., 2012; Cho et al., 2011; Halas et al., 2011) and high antimicrobial activity against pathogenic bacteria (Franz et al., 2010; Olafadehan et al., 2020).

In view of these abundant potentials in EOs, this study was designed to evaluate the effect of dietary supplementation of turmeric oil on the hematology and serum biochemical indices of broiler chickens. This experiment will further help to bridge the gap between food safety and livestock production

MATERIALS AND METHODS

Experimental Site

The experiment was carried out at the University of Abuja Teaching and Research Farm, Animal Science Section, Main Campus, along Airport Road, Gwagwalada, Abuja, Nigeria.

Extraction of turmeric oil (TOL)

Fresh turmeric rhizome was purchased from an open market in Gwagwalada, Abuja. The outer layer of the rhizome was peeled using a kitchen knife, it was thereafter separated and sun-dried for one week. The dried rhizome was granulated into coarse particles using a laboratory grinder. A 100 g of grinded rhizome placed onto a thimble and the thimble was put into the sohxlet extractor. N-hexane solvent was poured into three-neck- round bottom flask that is joined with the extractor and flask along with the condenser on the top to avoid any solvent losses. The whole assembly was then placed on the temperature controller heater to provide the required

temperature. The temperature was measured by a thermometer that was inserted in one of the necks of the round bottom flask. After certain interval of the time the experiment was stopped and the trapped oil in the solvent was separated. The mixture of solvent and oil was separated using rotary evaporator under vacuum at temperature of 65oC, the oil obtained after evaporation was weighed.

Pre-experimental operations

Prior to the commencement of the experiment, pens were properly disinfected, drinkers and feeder were properly washed, electrical fittings were fixed and foot dip was provided to ensure proper biosecurity.

Management of experimental animals

200- 1- day old (Ross 308) broiler chicks with mixed sex were used for the experiment. The birds were purchased from a commercial hatchery in Ibadan, Nigeria. It was weighed on arrival to obtain their initial body weight and thereafter weekly. A deep litter housing system was used and birds were divided to five treatments with four replicates consisting of 10 birds in a completely randomized design. Electric bulbs (200 Watts) were used as heat source, complimented with charcoal pot to maintain temperature, wood shavings served as the litter material. Vaccination was done according to the prevailing disease condition in the environment and all other management practices were strictly adhered to throughout the experiment which lasted for 56 days

Ration formulation

The basal diet was formulated to meet the nutrient requirements of birds according to NRC (1994) as presented in Table 1.

Treatment 1: Basal diet + 0 % TOL

Treatment 2: Basal diet + 0.1 % TOL

Treatment 3: Basal diet + 0.2 % TOL

Treatment 4: Basal diet + 0.3 % TOL

Treatment 5: Basal diet + 0.4 % TOL

DATA COLLECTED

Chemical analysis

Proximate compositions of experiment diet were determined by using official method of analysis by AOAC (2000).

Hematological and serum biochemical analysis

Blood samples were collected very early in the morning from the wing vein from three (3) randomly selected birds per replicate into a 5 ml sterile syringe using 23-gauge needles and transferred into an ethylene diamine tetra acetic acid (EDTA) bottle. Hematological parameters: pack cell volume (PCV), red blood cell (RBC), hemoglobin (Hb), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), mean corpuscular volume (MCV), white blood cell (WBC) and its differentials were analyzed using an automated machine (Sysmex, Model KU-30 HG, India).

Serum analysis was carried out using bottles free from EDTA, blood was analyzed for total protein, albumin, globulin, cholesterol, alanine transaminase (ALT) and aspartate transaminase (AST) were assayed using diagnostic kit manufactured by Merck India Ltd (Model PS-09R)

Statistical analysis

All data were subjected to one -way analysis of variance (ANOVA) using SPSS (23.0) and significant means were separated using Duncan multiple range tests (Duncan, 1955). Significant was declared if $P \leq 0.05$.

RESULTS AND DISCUSSIONS

Table 1: Percentage composition of experimental diet

Ingredients	Starter mash (0-4 weeks)	Finisher mash (5-8 weeks)
Maize	52.00	60.00
Soya meal	38.60	30.10
Groundnut cake	3.00	4.00
Fish meal (72%)	1.00	-
Bone meal	3.00	3.00
Limestone	1.50	2.00
Lysine	0.15	0.15
Methionine	0.20	0.20
Toxin binder	0.01	0.01
*Premix	0.25	0.25
Salt	0.30	0.30
Total	100.00	100.00

Determined analysis (% DM)

Crude protein	23.23	20.91
Crude fibre	3.14	4.00
Ether extract	5.01	4.74
Calcium	1.28	1.31
Phosphorus	0.63	0.68
Energy	2901.9	3100.7

* Premix supplied per kg diet :- Vit A, 10,000 I.U; Vit E, 5mg; Vit D3, 3000I.U, Vit K, 3mg; Vit B2, 5.5mg; Niacin, 25mg ; Vit B12, 16mg ; Choline chloride, 120mg ; Mn, 5.2mg ; Zn, 25mg ; Cu, 2.6g ; Folic acid, 2mg ; Fe, 5g ; Pantothenic acid, 10mg ; Biotin, 30.5g ; Antioxidant, 56mg.

Hematological parameters of broiler chicken fed diet supplemented with turmeric oil

Table 2 reveals the hematological parameters of broiler chicken fed turmeric oil (TOL). PCV, Hb, RBC, MCV, MCH and MCHC ranged between 30.10 – 38.96 %, 9.88 – 13.80 g/dl, 2.01 – 3.95 ($10^6/\mu\text{L}$), 101.2 – 149.8 fl, 32.77 – 49.15 pg and 30.49 – 35.53 % respectively. Highest value was recorded in T4 and T5, intermediate in T2, T3 and lowest in T1 ($P < 0.05$). The PCV, Hb, RBC and WBC values increased as the level of turmeric oil increases in the diets of the animals ($P < 0.05$). This result is in agreement with the findings of Gerardo *et al.* (2017) who noted that administration of Mexican oregano oil at 0.4g/kg showed positive haematological activities in broiler chicks. Similar recommendation was made by Alagbe and Grace (2019); Hippenstiel *et al.* (2014); Cubuk *et al.* (2006) who reported a significant difference ($P < 0.05$) in broilers fed herbal essential oil as a Phyto additive. This effect could be attributed to the presence of bioactive chemicals or secondary metabolites in turmeric oil. However, all the values of the haematological parameters fall within the ranges for broilers as reported by Talebi *et al.* (2005); Ibrahim Albokhadaim (2012). Hematological studies have been found useful for disease prognosis and for therapeutic and stress monitoring (Braun *et al.*, 2010) but can vary due to age, gender, environment, infection and poisoning (Khan and Zafar, 2005; Abdi-Hachesoo *et al.*, 2011). Red blood cell is involved in the transport of oxygen and carbon dioxide in the body (Isaac *et al.*, 2013). This is a clear indication that birds in T5 will have a have enough oxygen especially in situation of oxygen starvation. Nse Abasi *et al.* (2014); Subhadarsini and Silpa, 2020 reported that hematocrit or PCV is an index of toxicity; lower value could be a sign of anemia.

WBC, leucocytes, monocytes, heterophils, basophils and eosinophils values ranged between 19.89 – 30.22 ($10^3/\mu\text{L}$), 14.18 – 18.22 %, 0.99 – 1.44 %, 5.09 – 7.58 %, 1.04 – 2.90 % and 1.03 – 1.74 % respectively. Significant differences ($P<0.05$) were observed among the treatments, WBC helps to fight against infections and provide resistance against diseases (Soetan *et al*, 2013). This is an indication that birds in T4 and T5 have high resistance to infections which amounts to low mortality and healthy stocks. Basophils and eosinophils play a role in regulating allergic and inflammatory processes and host defense responses against parasitic infections like helminthiasis and ectoparasitic infestation (Butterworth, 1999; Alagbe *et al.*, 2020).

Table 2: Haematological parameters of broiler chicken fed diet supplemented with turmeric oil

Parameters	T1	T2	T3	T4	T5	SEM
PCV (%)	30.10 ^c	35.43 ^b	35.56 ^b	38.71 ^a	38.96 ^a	0.03
Hb (g/dl)	9.88 ^c	11.01 ^c	12.06 ^b	13.71 ^b	13.80 ^a	0.09
RBC $\times 10^6/\mu\text{L}$	2.01 ^c	3.61 ^b	3.68 ^b	3.71 ^a	3.95 ^a	0.10
MCV (fl)	149.8 ^a	103.7 ^c	107.5 ^b	107.0 ^b	101.2 ^c	0.18
MCH (pg)	49.15 ^a	30.50 ^c	32.77 ^c	36.95 ^b	34.94 ^b	2.33
MCHC (%)	32.82 ^c	29.41 ^b	30.49 ^b	35.53 ^a	33.51 ^a	3.09
WBC $\times 10^3/\mu\text{L}$	19.89 ^c	22.56 ^b	25.35 ^b	28.05 ^a	30.22 ^a	1.25
Lymphocytes %	14.18 ^c	15.90 ^c	16.56 ^b	17.08 ^a	18.22 ^a	0.21
Monocytes %	0.99 ^c	1.44 ^b	1.33 ^b	1.35 ^b	1.38 ^a	0.07
Heterophils %	5.09 ^c	6.06 ^b	6.17 ^c	6.45 ^b	7.58 ^a	0.12
Basophils %	1.04 ^c	2.10 ^c	2.17 ^b	2.23 ^b	2.90 ^a	0.09
Eosinophils %	1.03 ^c	1.41 ^b	1.51 ^a	1.58 ^a	1.74 ^a	0.01

^{abc} means different superscript along rows differs significantly at $P<0.05$

PCV: pack cell volume; Hb: hemoglobin; MCV: mean corpuscular volume; MCH: mean corpuscular hemoglobin; MCHC: mean corpuscular hemoglobin concentration.

Serum biochemical indices of broiler chicks fed diets supplemented with turmeric oil (TOL)

Table 3 reveals the serum biochemical indices of broiler chicks fed diets supplemented with TOL. The total protein, albumin, globulin, cholesterol, ALT and AST values ranged between 3.45 – 4.95 g/dl, 1.83 – 2.95 g/dl, 1.62 – 2.00 g/dl, 35.1 – 78.1 mg/l, 48.10 – 83.19 iu/L and 85.10 – 103.2 iu/L respectively. All the values were significantly affected ($P<0.05$) by the dietary supplementation of TOL. The total protein in the serum of animals are influenced by protein quality in the diet, this showed that the protein levels in the diet was enough to support normal protein reserves across the treatments (Olabanji *et al.*, 2007; Alagbe, 2020; Adewale *et al.*, 2021). However, the values obtained in this study were within the normal ranges for broilers reported by Livingstone *et al.* (2020); Obiakaonu *et al.* (2011). Cholesterol level decreased as the level of turmeric increased in the diet of the animal, this is a clear indication that TOL could perform hypolipidemic activity, thus preventing cardiovascular infection (Alagbe, 2021). Alanine transaminase (ALT) and aspartate transaminase were depressed as the level of TOL increased indicating no toxicity (Musa *et al.*, 2020; Oluwafemi *et al.*, 2020).

Table 3: Serum biochemical indices of broiler chicks fed diets supplemented with TOL

Parameters	T1	T2	T3	T4	T5	SEM
Total protein (g/dl)	3.45 ^b	4.61 ^a	4.70 ^a	4.65 ^a	4.95 ^a	0.98
Albumin (g/dl)	1.83 ^b	2.87 ^a	2.70 ^a	2.85 ^a	2.95 ^a	0.02
Globulin (g/dl)	1.62 ^b	1.74 ^b	2.00 ^a	2.00 ^a	2.00 ^a	0.31
Cholesterol (mg/l)	78.1 ^a	48.3 ^b	45.1 ^b	38.3 ^b	35.1 ^b	2.33
ALT (iu/L)	83.19 ^a	50.61 ^b	50.10 ^b	49.41 ^b	48.10 ^b	4.12
AST (iu/L)	103.2 ^a	97.40 ^b	90.31 ^b	88.17 ^c	85.10 ^c	2.78

^{abc} means different superscript along rows differs significantly at $P<0.05$

ALT: Alanine transaminase; AST: Aspartate transaminase

CONCLUSION

It was concluded that turmeric oil has a great potential and can be generally considered natural, less toxic due to the presence of various secondary metabolites and can be supplemented in the diet of broilers up to 0.4 % without causing any deleterious effect on the health and general performance of broiler chicks.

REFERENCES

1. Association of Official Analytical Chemistry (2000). Official Method of Analysis, 15th edition. Washington D.C., USA.
2. NRC (1994). Nutrient Requirements of Swine (11th ed). National Research Council, National Academy of Science. National Academy Press, Washington, DC.
3. Talebi, A., Asri-Rezaei, S., Rozeh-Chai, R and Sahraei, R. (2005). Comparative studies on haematological values of broiler strains (Ross, Cobb, Arbo-acres and Arian). International Journal of Poultry Science, 4(8):573-579.
4. Livingston, M.L., Cowieson, A.J., Crespo, R., Hoang, V., Nogal, B and Browning, M. (2020). Effect of broiler genetics, age and gender on performance and blood chemistry. Heliyon 6 (2020) e 04400.
5. Abdi-Hachesoo, B., Talebi, A and Asri-Rezaei, S. (2011). Comparative study on blood profile of indigenous and Ross-308 broiler breeders. Global Veterinary Journal, 7:238-241.
6. Alagbe, J.O. (2020). Performance, hematology and serum biochemical parameters of weaner rabbits fed different levels of fermented *Lagenaria breviflora* whole fruit extract. Advances in Research and Reviews, 2020, 1:5.
7. Obiakaonu, H.O., Okoli, I.C., Opara, M.N., Okoro, V.M.O., Ogbuewu, I.P., Etuk, E.B and Udedibie. A.B.I (2011). Haematological and serum biochemical indices of starter broilers fed neem leaf meal. Online Journal of Animal and Feed Research, 1(4):150-154.
8. Subhadarsini, M and Silpa, M.G. (2020). Comparative haematology and biochemical parameters of Indigenous broiler chicken. International Journal of Scientific Technology Research, 9(4):972-978.
9. Singh, A.S., Alagbe, J.O., Sharma, S., Oluwafemi, R.A and Agubosi, O.C.P. (2021). Effect of dietary supplementation of melon (*Citrullus lanatus*) seed oil on the growth performance and antioxidant status of growing rabbits. International Journal of Orange Technologies 3(3): 19-30.
10. Alagbe, J.O (2021). Daniellia oliveri leaf extracts as an alternative to antibiotic feed additives in broiler chicken diets: meat quality and fatty acid composition. International Journal of Clinical and Medical Informatics 4(1): 15-24.
11. Adewale, A.O., Alagbe, J.O and Adeoye, Adekemi Grace (2021). Dietary Supplementation of Rauvolfia Vomitoria Root Extract as a Phytogenic Feed Additive in Growing Rabbit Diets: Haematology and Serum Biochemical Indices. Scopia International Journal for Science, Commerce and Arts 1(3): 16-27.
12. Oluwafemi, R.A., Oluwayinka, E.O and Alagbe, J.O. (2020). Effect of dietary supplementation of neem oil (*Azadirachta indica*) on the growth performance and nutrient digestibility of weaned rabbits. European Journal of Biotechnology and Bioscience. 8(5): 6-10.
13. Musa, B., Alagbe, J.O., Adegbite Motunrade Betty, Omokore, E.A. (2020). Growth performance, caeca microbial population and immune response of broiler chicks fed aqueous extract of *Balanites aegyptiaca* and *Alchornea cordifolia* stem bark mixture. United Journal for Research and Technology, 2(2):13-21
14. Alagbe, J.O and Grace, F.R. (2019). Effect of *Albizia lebeck* seed oil dietary supplementation on the haematological and serum biochemical parameters of weaner rabbits. Sumerianz Journal of Agriculture and Veterinary. 2(10): 96 -100.

15. Alagbe, J.O., Sharma, D and Xing Liu (2019). Effect of aqueous *Piliostigma thonningii* leaf extracts on the haematological and serum biochemical indices of broiler chicken. *Noble International Journal of Agriculture and Food Technology*. 1(2): 62-69.
16. Alagbe, J.O. (2019). Haematology, serum biochemistry, relative organ weight and bacteria count of broiler chicken given different levels of *Luffa aegyptiaca* leaf extracts. *International Journal of Advanced Biological and Biomedical Research*. 7(4):382-392.
17. Nse Abasi, N.E., Mary, E.W., Uduak, A and Edem, E.A.O (2014). Haematological parameters and factors affecting their values. *Journal of Agricultural Science*, 2(1): 37-47.
18. Isaac, L. J., Abah, G., Akpan, B and Ekaette, I.U (2013). Haematological properties of different breeds and sexes of rabbits. *Proceedings of the 18th Annual Conference of Animal Science Association of Nigeria*. (Pg. 24-27).
19. Olabanji, R.O., GO Farinu., JA Akinlade., OO Ojebiyi., AA Odunsi and AA Akingbade (2007). Studies on Haematological and Serum Biochemical Characteristics of Weaner Rabbits Fed Different Levels of Wild Sunflower (*Tithonia diversifolia* Hems. A. Gray) Leaf- Blood Meal Mixture. *International Journal of Agriculture and Apiculture Research*. 4 (1&2): 80-89.
20. Alagbe, J.O., Agubosi, O.C.P., Ajagbe, A.D, Shittu, M.D and Akintayo Balogun, O.M (2020). Performance, haematology and serum biochemical parameters of growing grass cutters fed *Phyllanthus amarus* and *Piliostigma thonningii* leaf meal mixture as partial replacement for Soya bean meal. *United International Journal for Research and Technology*, 2(1): 14-23.
21. Mahima, A., Rahal, R, Deb, S.K., Latheef, H.A. (2012). Immunomodulatory and therapeutic potentials of herbal, traditional and ethnoveterinary medicines. *Pakistan Journal of Biological Sciences*, 15:754-774.
22. Oluwafemi, R.A., Egwuiyi. G.N and Alagbe, J.O. (2020). Effect of feeding *Polyalthia longifolia* leaf meal as partial replacement of wheat offal. *European Journal of Agricultural and Rural Education*. 1(1), 8-16.
23. Alagbe, J.O (2021). *Daniellia oliveri* leaf extracts as an alternative to antibiotic feed additives in broiler chicken diets: meat quality and fatty acid composition. *International Journal of Clinical and Medical Informatics* 4(1): 15-24.
24. Alagbe, J.O and Akintayo-Balogun, O.M. (2020). Effects of dietary supplementation of *Albizia lebbek* seed oil (ALO) on the fatty acid composition of weaner rabbits. *Biochemistry and Biotechnology Research*, 8(2): 29-33.
25. Oluwafemi, R.A., Akinbisola, S.A and Alagbe, J.O. (2020). Nutritional and growth performance of feeding *Polyalthia longifolia* Leaf Meal as partial replacement of Wheat Offal in the diet of broiler chicks. *European Journal of Biotechnology and Bioscience*. 8(4): 17-21.
26. Oluwafemi, R.A., Isiaka Olawale and Alagbe, J.O. (2020). Recent trends in the utilization of medicinal plants as growth promoters in poultry nutrition- A review. *Research in: Agricultural and Veterinary Sciences*. 4(1): 5-11.
27. Olafadehan, O.A., Oluwafemi, R.A and Alagbe, J.O. (2020). Carcass quality, nutrient retention and caeca microbial population of broiler chicks administered Rolfe (*Daniellia oliveri*) leaf extract as an antibiotic alternative. *Journal of Drug Discovery*. 14(33):146-154.
28. Bouhaddouda N, Aouadi S, Labiod R. Evaluation of chemical composition and biological activities of essential oil and methanolic extract of *Origanum vulgare* L. ssp. *glandulosum* (Desf.) Ietswaart from Algeria (2016). *Int J Pharmacognosy Phytochem Res*.8:104e12.
29. Franz C, Baser K, Windisch W. Essential oils and aromatic plants in animal feeding-a European perspective (2010). A review. *Flavour Fragr* 25:327e40.
30. Franz C, Novak J. Sources of essential oils. In: Baser KHC, Buchbauer G, editors (2009). *Handbook of essential oils: science, technology, and applications*. Boca Raton: CRC Press/Taylor & Francis Group; p. 39e82.
31. Giannenas I, Bonos E, Christaki E, Florou-Paneri P. Essential oils and their applications in animal nutrition (2013). *Med Aromat Plants* 2:1e12.
32. Hammer KA, Carson CF. Antibacterial and antifungal activities of essential oils. In: Thormar H, editor. *Lipids and essential oils as antimicrobial agents* (2011). Iceland: Wiley; p. 255e306.

33. Kommera SK, Mateo RD, Neher FJ, Kim SW. Phytobiotics and organic acids as potential alternatives to the use of antibiotics in nursery pig diets (2006). *Asian-Australas J Anim Sci* 19:1784e9.
34. Kim J, Marshall MR, Wei C. Antibacterial activity of some essential oil components against five foodborne pathogens (1995). *J Agric Food Chem* 43:2839e45.
35. Oke F, Aslim B, Ozturk S, Altundag S. Essential oil composition, antimicrobial and antioxidant activities of *Satureja cuneifolia* ten (2009). *Food Chem* 112:874e9.
36. Purchiaroni F, Tortora A, Gabrielli M, Bertucci F, Gigante G, Ianiro G, et al. The role of intestinal microbiota and the immune system (2013). *Eur J Rev Med Pharmacol Sci* 17:323e33.
37. Wenk C. Herbs and botanicals as feed additives in monogastric animals (2003). *Asian- Australas J Anim Sci* 16:282e9.
38. Zhang Y, Gong J, Yu H, Guo Q, Defelice C, Hernandez M, et al. Alginate-whey protein dry powder optimized for target delivery of essential oils to the intestine of chickens (2014). *Poult Sci* 93:2514e25.

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